

**SAFETY Meeting Minutes**  
UAMS IBC

**MEETING TIME RECORDS**

**Meeting start time:** 11/7/2025 12:01 PM

**Meeting end time:** 11/7/2025 12:18 PM

**Meeting type:** Virtual

<b>Name of Regular/Alternate Member</b>	<b>Status (Member or Alternate)</b>	<b>Present by Teleconference?</b>
Ha-Neui Kim	Member	Yes
Matthew Jorgenson	Member	Yes
Robert Hunter	Member	No; electronic vote
Kimberly Murphy	Member	No
Lindsey Clark	Member	Yes
James Douglas	Member	No
Amanda Holloway	Ex Officio	No
James Bishop	Member	No
Youssef Aachoui	Member	No
Jia Liu	Member	Yes
Yuet-Kin Leung	Member	Yes
Melaney Gee	Member	Yes
Mark Manzano	Member	Yes
Christine Simecka Morgan	Member	No
Antino Allen	Member	Yes
KyoungHyun Kim	Member	Yes
James Townsend	Ex Officio	No
Shengyu Mu	Member	No
Kikumi Ono-Moore	Ex Officio	No
Zhiqiang Qin	Member	Yes

**QUORUM INFORMATION**

**Number of SAFETY members on the roster:** 17

**Number required for quorum:** 9

**Quorum:** Present

All members present via teleconference received all pertinent material before the meeting and were able to actively and equally participate in all discussions.

ATTENDANCE STATUS AND VOTING KEY	
ABSTAIN:	Present for the vote but not voting “For” or “Against.”
ABSENT:	Absent for discussion and voting for reasons other than a conflict of interest.
RECUSED:	Absent from the meeting during discussion and voting because of a conflict of interest.
SUBSTITUTION:	When regular members and their alternate(s) are listed in the ATTENDANCE table above and an alternate member serves as a substitute for the regular member this identifies the name of the alternate to indicate which individual is serving as the voting member for this vote. May be deleted if there are no substitutions.

GUEST NAMES
N/A

**Previous Meeting minutes approved:** Yes

## REVIEW OF SUBMISSIONS

The review and discussion of the protocols listed below included the following elements: the agents involved and their characteristics; types of manipulations planned; the source(s) and nature of the nucleic acid sequences; the host organism(s) and vector(s) to be utilized; whether expression of a foreign gene is intended and, if so, the specific protein(s) to be produced; the containment conditions to be applied, including biosafety level and any special provisions; and the relevant sections of the NIH Guidelines.

All IBC members present were reminded to identify any conflicts of interest as each registration was reviewed.

For each protocol reviewed, it was confirmed that the Principal Investigator (PI) and laboratory personnel have received appropriate training in the safe conduct of research.

**Initial Protocol****1. Review of SPROTO202400000041**

Title:	Cell line engineering
Investigator:	<a href="#">Tudor Moldoveanu</a>
Submission ID:	SPROTO202400000041
Description:	<p>Research in my lab seeks to investigate structurally and functionally program cell death pathways and to probe these pathways with chemical biology tools.</p> <p>The primary objective for the IBC is to enable production of cell lines of certain genetic background by knock out and/or over-expression of cell death pathway components. We primarily use retrovirus expression in mammalian cells and baculovirus expression in insect cells to manipulate components of the cell death pathways. In some occasions, when retroviral expression is poorly behaved, we may use lentivirus or the bacmam system (baculovirus infection of mammalian cells) to express our proteins of interest.</p>
Agent Containment:	<p>Biological Containment Levels:</p> <ul style="list-style-type: none"> <li>• Retrovirus: BSL-2</li> <li>• HCT116: BSL-2</li> <li>• MV4-11 (Human Acute Myeloid Leukemia Cell Line): BSL-2</li> <li>• MOLM-13 (Human Acute Myeloid Leukemia Cell Line): BSL-2</li> <li>• HEK293T Human Cell Line: BSL-2</li> <li>• E. coli: BSL-1</li> <li>• MEF (Mouse Embryo Fibroblasts): BSL-2</li> <li>• HeLa cells: BSL-2</li> </ul>
Applicable NIH Guidelines:	<ul style="list-style-type: none"> <li>• Section III-D-1-a</li> <li>• Section III-F-3</li> <li>• Section III-E</li> <li>• Section III-F-8-C-II</li> <li>• Section III-D-1</li> <li>• Section III-D</li> <li>• Section III-E-1</li> </ul>

a. **Determination:** Modifications Required

b. **Required modifications:**

Minor revisions necessary. Please review and respond to all comments throughout the protocol.

Contact BSOs with any questions. Thank you!

c. **Votes:**

**For:** 11

**Against:** 0  
**Recused:** 0  
**Absent:** 6  
**Abstained:** 0

## Initial Protocol

### 2. Review of SPROTO202500000050

Title:	Oncolytic Immunotherapy for cancers
Investigator:	<a href="#">Mulu Tesfay</a>
Submission ID:	SPROTO202500000050
Description:	<p>RNA viruses such as vesicular stomatitis virus (VSV), a prototype for negative-stranded RNA viruses, have been commonly used as an oncolytic virus to treat different types of cancers due to their ability to selectively kill cancer cells while sparing surrounding normal cells. Furthermore, VSV grows to very high titers in most animal cells and can be prepared in large quantities. Recombinant VSV is currently being tested in many clinical trials involving human cancer patients. Our lab is interested in using VSV and VSV-derived recombinant viruses to infect and selectively kill cancer cells of the liver and pancreas. Our lab plans to characterize the VSV-recombinant viruses in vitro and in vivo settings to ascertain their proposed efficacy capabilities and safety profiles. We plan to characterize these viruses for their oncolytic and safety properties in various solid tumors, including liver, pancreatic, and eye cancers (an eye cancer that metastasizes to the liver), both in vitro and in vivo. We will test ways to enhance the oncolytic potential of VSV-recombinant viruses and explore the potential for combination therapy with other biologics, small molecules (such as chemotherapy) and checkpoint inhibitors (ICIs). We also plan to characterize existing, approved, live-attenuated vaccines, such as MMR and 17D, for their immunomodulatory potential in solid tumors.</p>
Agent Containment:	<p>Biological Containment Levels:</p> <ul style="list-style-type: none"> <li>• PANC1: BSL-2</li> <li>• B16-F10: BSL-2</li> <li>• RILWT Cell Line: BSL-2</li> <li>• Animal Tissue: BSL-2</li> <li>• A549: BSL-2</li> <li>• MP41 human tumor cell line: BSL-2</li> <li>• B16LS9 Murine cutaneous melanoma cells: BSL-2</li> <li>• BxPC-3 CRL-1687 Human Cell Line: BSL-2</li> </ul>

	<ul style="list-style-type: none"> <li>• RIL-175: BSL-2</li> <li>• 92.1 human tumor cell line: BSL-2</li> <li>• CT26: BSL-2</li> <li>• Salmonella: BSL-2</li> <li>• Vaccinia virus: BSL-2</li> <li>• Vescicular Stomatitis Virus: BSL-2</li> <li>• 293T: BSL-2</li> <li>• Jurona Virus (JURV) Attenuated: BSL-2</li> <li>• KPCY Mouse Pancreatic Cancer Cell Line (7160c2): BSL-2</li> <li>• BHK-21: BSL-2</li> <li>• EMT6: BSL-2</li> <li>• MP46 human tumor cell line: BSL-2</li> <li>• HeLa cells: BSL-2</li> <li>• Pan02: BSL-2</li> <li>• Animal Blood: BSL-2</li> <li>• Hep3B Human Cell Line: BSL-2</li> <li>• Vero: BSL-2</li> <li>• E. coli: BSL-2</li> <li>• Hepa 1-6: BSL-2</li> </ul>
Applicable NIH Guidelines:	<ul style="list-style-type: none"> <li>• Section III-D-1-a</li> <li>• Section III-D-1</li> <li>• Section III-D</li> </ul>

a. **Determination:** Modifications Required

b. **Required modifications:**

Minor revisions necessary. Please review and respond to all comments throughout the protocol.

Contact BSOs with any questions. Thank you!

c. **Votes:**

<b>For:</b>	11
<b>Against:</b>	0
<b>Recused:</b>	0
<b>Absent:</b>	6
<b>Abstained:</b>	0

## De Novo Review

### 3. Review of SPROTO202500000074

Title:	Immunity to Human Papillomavirus (BP144)
Investigator:	<a href="#">Mayumi Nakagawa</a>
Submission ID:	SPROTO202500000074
Description:	Human papillomavirus (HPV) is best known for being the

	<p>causative agent of cervical cancer, which is the fourth most common cancer among women globally. It can also cause vaginal, vulvar, penile, oropharyngeal, and anal cancers and is estimated to be responsible for 5.2% of cancer burden in the world.</p> <p>To date, more than 120 types of HPV have been described. The ones that have genital tropism and are associated with the development of malignancy are called high-risk, and they include HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 53, 56, 58, 59, 66, 67, 68, 69, 70, 73, and 82. Low-risk HPV types also have genital tropism and are associated with development of genital warts. They include HPV types 6, 11, 40, 42, 54, 61, 62, 71, 72, 81, 83, 84, and CP6108. Some HPV types have cutaneous tropism and are associated with common warts. These most commonly include HPV types 2, 27, and 57. Immunity is known to play a key role in affecting the course of HPV pathogenesis. Furthermore, enhancement of anti-HPV immunity through vaccines and immunotherapies are promising means to prevent and treat conditions associated with HPV. However, it is still not easy to grow a sufficient quantity of HPV to be used as a reagent in immune assay to measure anti-HPV T-cell responses. Therefore, scientists rely on recombinant techniques to produce HPV antigens.</p> <p>Recombinant vaccinia viruses expressing viral genes such as E1/E4, E2, E5, E6, E7, L1, or L2 are used to infect cells to present endogenously processed epitopes to CD8 T-cells which have cytotoxic capability, and can lyse infected cells. Recombinant fusion proteins (such as ones co-expressed with glutathione S-transferase) are pulsed with antigen-presenting cells to process epitopes to be presented to CD4 T-cells which have helper functions.</p> <p>We already have recombinant vaccinia viruses that express 16 E6, 16 E7, 16 L1, 16 L2, 18 E6, 18 E7, 35 E6, 52 E6, and 57 L1, and recombinant fusion proteins for 16 E1/E4, 16 E5, 16 E6, 16 E7, 18 E6, 35 E6, and 52 E6 proteins.</p> <p>No additional cloning experiments are planned.</p> <p>The renewal is for continued use of vaccinia viruses expressing 16 E6, 16 E7, 18E6, 35 E6, and 52 E6 and recombinant GST-fusion proteins expressing 16 E6, 16 E7, 18 E6, 35 E6, and 52 E6. They are being cultured with antigen presenting cells such as dendritic cells, and are used for in</p>
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	<p>vitro stimulation of T cell lines. In turn, the T cells are used in immune assay such as IFN-g enzyme-linked immunospot assay to assess anti-HPV immune responses.</p> <p>All samples are being handled in BSCs at UAMS. The biosafety concern is for use of vaccinia virus in primary human blood cells. There will be no genetic manipulations of primary human blood cells. We culture human blood cells for 3 weeks, they are not immortalized, so they are not considered to be cell lines. We are using CV-1 cells. The relevant NIH guidelines include Section III-D and Section III-D-1. Risks are mitigated through training, use of universal precautions, and PPE. The risk group level is RG-2, biological research standards is BSL-2, and physical containment level is BL-2.</p> <p>Human stool and oral wash samples are also collected. DNA is extracted from human stool and oral wash samples, and are analyzed for microbiome composition.</p>
Agent Containment:	<p>Biological Containment Levels:</p> <ul style="list-style-type: none"> <li>• Human Blood: BSL-2</li> <li>• Vaccinia virus: BSL-2</li> <li>• CV-1 cells: BSL-2</li> <li>• Buccal Rinse: BSL-2</li> <li>• Human Stool: BSL-2</li> </ul>
Applicable NIH Guidelines:	<ul style="list-style-type: none"> <li>• Section III-D-1</li> <li>• Section III-D</li> </ul>

a. **Determination:** Modifications Required

b. **Required modifications:**

Minor revisions necessary. Please review and respond to all comments throughout the protocol.

Contact BSOs with any questions. Thank you!

c. **Votes:**

<b>For:</b>	11
<b>Against:</b>	0
<b>Recused:</b>	0
<b>Absent:</b>	6
<b>Abstained:</b>	0

## REVIEW OF OTHER AGENDA ITEMS

- Administrative approvals were acknowledged and approved.
- No safety incidents to report.
- No inspection findings to report.

- Committee members were updated on issues related to the new workflow in Muse and notified that implementation of the workflow has been paused pending a solution.